

## **Laboratory for Advanced Electron and Light Optical Methods**

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### **Transmission EM Embedment of Histological Slide Sections**

Technique Provided by Dr. E. Dalldorf (via Dr. J. Wright)

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1. Examine stained sections, select the cells you want to embed. Mark out a small area on the back of the glass slide with a marker.
2. Remove the cover slip with xylene.
3. Scrape away all of the tissue section except the small area you want for EM.
4. Place a drop of 1% osmium in 0.1 M phosphate buffer on the section in a petri dish for 30 min under the fume hood.
5. Dehydrate the sections as per SOP 6.0.0. Put large drops of dehydration agents over the section(s).
6. Cut off the bottom of a BEEM embedding capsule. Place it over the dehydrated sections. Fill capsule with Spurr resin. Change after 1 hour.
7. Put in oven at 70° C for 1-2 hours. When it is firm but not hard, pry the BEEM capsule off of the slide.
8. Return the capsule to the oven and leave overnight to complete polymerization.
9. Trim block to tissue and cut sections.